



Myriad Morcells Fine™

is a conformable ECM graft
derived from 100% AROA ECM

AROA ECM
IS MORE THAN
JUST COLLAGEN

AROA ECM is extracellular matrix (ECM) technology that supplements patient's healing with important ECM proteins including growth factors, cytokines and anti-inflammatory proteins, known to help facilitate healing.^{1,2}

**MYRIAD MORCELLS FINE CONTAINS COLLAGEN AND
MANY ECM PROTEINS THAT ARE IMPORTANT IN HEALING³**

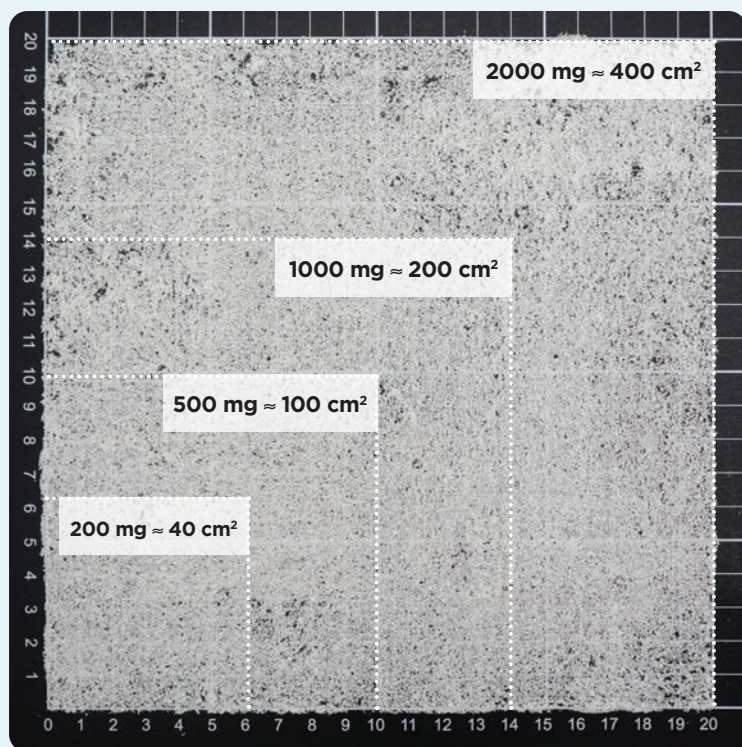
Product	Type of technology	Collagen type I	Collagen type II	Collagen type III	Collagen type IV	Fibronectin	Elastin	Hyaluronic acid	Growth factors and cytokines	Basement Membrane	Residual Vascular Channels
Myriad Morcells Fine	ECM ⁴	✓ ^{2,3}	✓ ²	✓ ^{2,3}	✓ ^{2,3}	✓ ^{2,3}	✓ ^{2,3}	✓ ³	✓ ^{2,3}	✓ ^{2,3}	✓ ⁴
MicroMatrix®	ECM ^{5,6}	✓ ^{6,7}		✓ ⁶	✓ ^{6,7}	✓ ⁶	✓ ⁶		✓ ⁶	✓ ⁶	
CellerateRX®	Reconstituted collagen	✓ ⁸									
Integra® Flowable Wound Matrix	Reconstituted collagen and chondroitin sulphate	✓ ^{9,10}									
NIVIS®	Reconstituted collagen	✓ ¹¹		✓ ¹¹							

Myriad Morcells Fine™

MYRIAD MORCELLS FINE

Provides more coverage

Myriad Morcells Fine provides approximately 4 times the coverage compared to CellerateRX¹²



Approximate coverage of Myriad Morcells Fine

Product Size	Approximate dry coverage
200 mg	≤ 40 cm ²
500 mg	≤ 100 cm ²
1000 mg	≤ 200 cm ²
2000 mg	≤ 400 cm ²

Table above: maximum Myriad Morcells Fine coverage based on representative image (left).

The approximate coverage indicated in table is a guide only and should not supercede professional judgement.

Coverage will vary between defects based on defect characteristics.

Image left: representative image illustrating Myriad Morcells Fine coverage



THE IDEAL FOUNDATION

DERIVED FROM **OVINE FORESTOMACH**, AROA ECM PROVIDES THE IDEAL BALANCE OF BIOLOGY & STRUCTURE TO SUPPORT TISSUE REGENERATION

ARO A ECM FACILITATES:

- 01 Restoration of functional tissue¹³⁻¹⁵
- 02 Rapid volumetric fill and robust tissue formation¹³⁻¹⁶
- 03 Rapid establishment of blood supply to nourish regenerating tissue¹⁷



THE OPTIMAL SOURCE

THE **OVINE FORESTOMACH** IS A UNIQUE AND HIGHLY VASCULAR ORGAN THAT CONSTANTLY REGENERATES

- 01 Is highly vascularized as it's the primary means of nutrient absorption^{18,19}
- 02 Has a high rate of tissue turnover & remodeling²⁰
- 03 Rich in biology, including stem cells & signaling molecules²⁰

SUSTAINABLE TECHNOLOGY

FROM BY-PRODUCT
TO BIOTECH

An abundant source tissue derived from pasture-raised New Zealand sheep³



Myriad Morcells Fine™

Myriad Morcells Fine™

Now available in a finer particulate

Stock no.	Product Size
PW02NA0200	200 mg
PW02NA0500	500 mg
PW02NA1000	1000 mg
PW02NA2000	2000 mg



Myriad Morcells™

Available in a range of pack sizes

Stock no.	Product Size
PW01NA0200	200 mg
PW01NA0500	500 mg
PW01NA1000	1000 mg
PW01NA2000	2000 mg



1. Badyal, S.F., The extracellular matrix as a biologic scaffold material. *Biomaterials*, 2007;28(25): p.3587-93. 2. Dempsey, S.G., et al., Functional Insights from the Proteomic Inventory of Ovine Forestomach Matrix. *J Proteome Res*, 2019. 18(4):p.1657-1668. 3. Lun, S., et al., A functional extracellular matrix biomaterial derived from ovine forestomach. *Biomaterials*, 2010. 31(16):p.4517-29. 4. Smith, M.J., et al., Further structural characterization of ovine forestomach matrix and multi-layered extracellular matrix composites for soft tissue repair. *J Biomater Appl*, 2021;36(6):p.996-1010. 5. Gilbert, T.W., et al., Production and characterization of ECM powder: implications for tissue engineering applications. *Biomaterials*, 2005;26(12):p.1431-5. 6. Sadler, K., et al., Proteomic composition and immunomodulatory properties of urinary bladder matrix scaffolds in homeostasis and injury. *Semin Immunol*, 2017. 29: p. 14-23. 7. Bolland, F., et al., Development and characterisation of a full-thickness acellular porcine bladder matrix for tissue engineering. *Biomaterials*, 2007. 28(6): p. 1061-70. 8. Stafyla, V. et al., Effect of collagen powder on lymphorrhea after modified radical mastectomy. A randomized controlled trial. *Eur J Gynaecol Oncol*, 2011. 32(2): p. 185-7. 9. Hodgkinson, T. and A. Bayat, Ex vivo evaluation of acellular and cellular collagen-glycosaminoglycan flowable matrices. *Biomater*, 2015. 10(4): p. 041001. 10. Yannas, I.V., Tissue regeneration by use of collagen-glycosaminoglycan copolymers. *Clin Mater*, 1992. 9(3-4): p. 179-87. 11. Tela Bio website: <https://irtelabio.com/news-releases/news-release-details/tela-bio-announces-us-commercial-launch-nivistm-fibrillar> (accessed April 10 2024). 12. Data on file. 13. Cormican, M. T., N. J. Creel, B. A. Bosque, S. G. Dowling, P. P. Rideout and W. M. Vassy (2023). "Ovine Forestomach Matrix in the Surgical Management of Complex Volumetric Soft Tissue Defects: A Retrospective Pilot Case Series." *ePlasty* 23: e66. 14. Bosque, B. A., C. Frampton, A. E. Chaffin, G. A. Bohn, K. Woo, C. DeLeonardis, B. D. Lepow, M. M. Melin, T. Madu, S. G. Dowling and B. C. H. May (2021). "Retrospective real-world comparative effectiveness of ovine forestomach matrix and collagen/ORC in the treatment of diabetic foot ulcers." *Int Wound J* 2021 Aug 6 (Epub ahead of print). 15. Chaffin, A. E., S. G. Dowling, M. S. Kosyk and B. A. Bosque (2021). "Surgical reconstruction of pilonidal sinus disease with concomitant extracellular matrix graft placement: a case series." *J Wound Care* 30(Sup7): S28-S34. 16. Duplechain, A.B., et al., Soft Tissue Reconstruction With Ovine Forestomach Matrix After Wide Excision of Plantar Fibromatosis. *ePlasty*, 2023. 2023(23): p. e20. 17. Irvine, S. M., J. Cayzer, E. M. Todd, S. Lun, E. W. Floden, L. Negron, J. N. Fisher, S. G. Dempsey, A. Alexander, M. C. Hill, A. O'Rourke, S. P. Cunningham, C. Knight, P. F. Davis, B. R. Ward and B. C. H. May (2011). "Quantification of in vitro and in vivo angiogenesis stimulated by ovine forestomach matrix biomaterial." *Biomaterials* 32(27): 6351-6361. 18. Topps J, Kay et al (1968). Digestion of concentrate and of hay diets in the stomach and intestines of ruminants. *Br J Nutr* 22, 261-280. 19. Engelhardt W and Hales J (1977). Partition of capillary blood flow in rumen, reticulum, and omasum of sheep. *American Journal of Physiology-Endocrinology and Metabolism* 232(1): E53. 20. Baldwin, R. L.; McLeod, K. R.; Klotz, J. L.; Heitmann, R. N. Rumen development, intestinal growth and hepatic metabolism in the pre- and post weaning ruminant. *J. Dairy Sci.* 2004, 87 (Suppl. E), E55-E65.

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RX Only. Prior to use, be sure to read the entire Instructions For Use package insert supplied with the product.

For product questions, sampling needs, or detailed clinical questions concerning our products in the US,
please call 1-877-627-6224 or email customerservice@aroa.com

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